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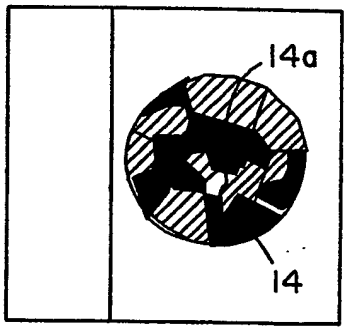

(57) Abstract

Disclosed are heterovesicular liposomes containing different biological compositions (14, 14a) each encapsulated in separate chambers of the liposomes, having defined size distribution, adjustable internal chamber size and number, methods of making them and treatment of patients with them. The preparation process includes the addition of composition to first lipid component in the vial to obtain an emulsion, the addition of composition to second lipid component in the vial to obtain a second emulsion and mixing these emulsions to form a chimeric emulsion.

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HETEROVESICULAR LIPOSOMES

Field of the Invention

The invention relates to the synthetic heterovesicular lipid vesicles or liposomes, processes for their manufacture and encapsulation of various materials therein, and treatment of patients with them.

Background Art

Multivesicular liposomes are one of the three main types of liposomes, first made by Kim, et al. (1983, Biochim, Biophys. Acta 782, 339-348), and are uniquely different from the unilamellar (Huang, 1969, Biochemistry 8,344-352; Kim, et al. 1981, Biochim. Biophys. Acta 646, 1-10) and multilamellar (Bangham, et al. 1965, J. Mol, Bio. 13,238-252) liposomes in that there are multiple non-concentric aqueous chambers within. Previously described techniques for producing liposomes relate to the production of non-multivesicular liposomes; for example, U.S. Patent Nos. 4,522,803 - Lenk, 4,310,506 - Baldeschwieler, 4,235,871 - Papahadjopoulos, 4,224,179 - 4,078,052 - Papahadjopoulos, 4,394,372 - Taylor, 4,308,166 - Marchetti, 4,485,054 - Mezei, and 4,508,703 - Redziniak. For a comprehensive review of various methods of liposome preparation, refer to Szoka, et al. 1980, Ann. Rev. Biophys. Bioeng. 9:467-508.

Heterovesicular liposomes are lipid vesicles or liposomes with multiple internal aqueous chambers where at least two substances of different compositions are each encapsulated in separate chambers within one liposomes. The lipid vesicles or liposomes with multiple internal aqueous

chambers include, but are not limited to, multilamellar liposomes, stable paucilamellar liposomes, and multivesicular liposomes. It is highly advantageous to provide a liposome delivery system in which two or more different substances are each encapsulated in separate compartments of a single liposome rather than encapsulated together in each compartment of the liposome.

Summary of the Invention

The composition of the present invention comprises heterovesicular liposomes, i.e. lipid vesicles or liposomes with multiple internal aqueous chambers where two or more substances of different compositions are each encapsulated separately in different chambers within one liposome.

Briefly, the method of the invention comprises making a "water-in-lipid" emulsion by dissolving amphipathic lipids in one or more organic solvents for the first lipid component, adding an immiscible first aqueous component including a substance to be encapsulated, preferably in the presence of hydrochloric acid, and then emulsifying the mixture mechanically. In the emulsion, the water droplets suspended in the organic solvent will form the internal aqueous chambers, and the monolayer of amphipathic lipids lining the aqueous chambers will become one leaflet of the bilayer membrane in the final product. A second lipid component is then formed by dissolving amphipathic lipids in a volatile organic solvent and adding an immiscible second aqueous component including a second substance to be encapsulated, preferably in the presence of hydrochloric acid. A second emulsion is then created. A chimeric emulsion is then formed by combining the first and second emulsions. The chimeric emulsion consists of multiple water droplets suspended in organic solvent where the substances of two different compositions are each dissolved separately in different aqueous droplets. The chimeric emulsion is then immersed in a third aqueous immiscible component preferably containing one or more nonionic osmotic agents

and acid-neutralizing agent of low ionic strength and then mechanically dividing it to form solvent spherules suspended in the third aqueous component. The solvent spherules contain multiple aqueous droplets where the substances of two different compositions are each dissolved separately in different aqueous droplets within a single solvent spherule. The volatile organic solvent is evaporated from the spherules preferably by passing a stream of gas over the suspension. When the solvent is completely evaporated, the spherules convert into heterovesicular liposomes with multiple internal aqueous chambers where two substances of different compositions are encapsulated separately in different chambers within one liposome.

The use of hydrochloric acid with a neutralizing agent, or other hydrochlorides which slow leakage rates is preferably for high encapsulation efficiency and for a slow leakage rate of encapsulated molecules in biological fluids and in vivo. It is also preferable to use neutralizing agent of low ionic strength to prevent solvent spherules from sticking to each other.

Accordingly, it is an object of the present invention to provide a heterovesicular lipid vesicle or liposome having at least two substances of different compositions each encapsulated separately in different chambers of the vesicle or liposome.

A further object of the present invention is the provision of a heterovesicular liposome containing at least two biologically active substances of different compositions each encapsulated separately in chambers of the liposome in the presence of hydrochloric acid or other hydrochlorides which slow the leakage of them.

It is a further object of the present invention to provide a heterovesicular liposome containing at least two biologically active substances of different compositions each encapsulated separately in chambers of the liposome in

the presence of hydrochloric acid or other hydrochlorides and a neutralizing agent.

It is a further object of the present invention to provide methods of producing such heterovesicular lipid vesicles or liposomes.

It is a further object of the present invention to provide processes for producing such heterovesicular lipid vesicles or liposomes by providing a first lipid component dissolved in one or more organic solvents and adding to the lipid component an immiscible first aqueous component containing a first substance to be encapsulated, forming a first water in oil emulsion from the first two immiscible components, providing a second lipid component dissolved in one or more organic solvents and adding into the lipid component an immiscible second aqueous component containing a second substance to be encapsulated, forming a second water in oil emulsion from the second two immiscible components, forming a chimeric emulsion by combining the first water in oil emulsion and second water in oil emulsion, transferring and immersing the chimeric emulsion into a third immiscible aqueous component, dispersing the chimeric emulsion to form solvent spherules containing multiple droplets of the first aqueous component containing the first substance and the second aqueous component containing the second substance, and evaporating the organic solvent from the solvent spherules to form the heterovesicular lipid vesicles or liposomes.

It is a further object to provide such a process in which a variety of hydrophilic biologically active materials and can be encapsulated separately in chambers of the heterovesicular lipid vesicles or liposomes.

It is a further object of the present invention to provide a method for the treatment of a patient with at least two separate biologically active substances of different compositions by administering them to the patient

encapsulated separately in chambers of a heterovesicular vesicle or liposome.

Other and further objects, features and advantages of the invention appear throughout the specification and claims.

Brief Description of the Drawings

Figures 1-8 are schematic diagrams illustrating preparation of a heterovesicular vesicle or liposome.

Description of Preferred Embodiments

The term "multivesicular liposomes" as used throughout the specification and claims means man-made, microscopic lipid-vesicles consisting of lipid bilayer membranes, enclosing multiple non-concentric aqueous chambers which all contain the same component. In contrast, the term "heterovesicular liposomes as used throughout the specification and claims means man-made, microscopic liquid vesicles consisting of lipid bilayer membranes enclosing multiple, aqueous chamber wherein at least two of the chambers separately contain substances of different compositions. The microscopic lipid vesicles include but are not limited to multilamellar liposomes, stable paucilamellar liposomes, and multivesicular liposomes.

The term "chimeric emulsion" as used throughout the specification and claims means an emulsion that consists of multiple water droplets suspended in organic solvent where the substances of two different compositions are each dissolved separately in different aqueous droplets.

The term "solvent spherule" as used throughout the specification and claims means a microscopic spheroid droplet of organic solvent, within which is multiple smaller droplets of aqueous solution. The solvent spherules are suspended and totally immersed in a second aqueous solution.

The term "neutral lipid" means oil or fats that have no membrane-forming capability by themselves and lack a hydrophilic "head" group.

The term amphipathic lipids means those molecules that have a hydrophilic "head" group and hydrophobic "tail" group and have membrane-forming capability.

The composition of the present invention is a heterovesicular lipid vesicle or liposome having at least two substances of different compositions each encapsulated separately in different chambers of the vesicle or liposome.

Many and varied biological substances can be incorporated by encapsulation within the multivesicular liposomes. These include drugs, and other kinds of materials, such as DNA, RNA, proteins of various types, protein hormones produced by recombinant DNA technology effective in humans, hematopoietic growth factors, monokines, lymphokines, tumor necrosis factor, inhibin, tumor growth factor alpha and beta, mullerian inhibitory substance, nerve growth factor, fibroblast growth factor, platelet-derived growth factor, pituitary and hypophyseal hormones including LH and other releasing hormones, calcitonin, proteins that serve as immunogens for vaccination, and DNA and RNA sequences.

The following Table 1 includes a list of representative biologically active substances which can be encapsulated in heterovesicular liposomes in the presence of a hydrochloride and which are effective in humans.

TABLE 1

Antiasthma	Antiarrhythmic	Tranquilizers
metaproterenol	propanolol	chlorpromazine
aminophylline	atenolol	benzodiazepine
theophylline	verapamil	butyrophenones
terbutaline	captopril	hydroxyzines
Tegretol	isosorbide	meprobamate
ephedrine		phenothiazines
isoproterenol		reserpine
adrenalin		thioxanthines
norepinephrine		

Cardiac glycosides
digitalis
digitoxin
lanatoside C
digoxin

Hormones
antidiuretic
corticosteroids
testosterone
estrogen
thyroid
growth
ACTH
progesterone
gonadotropin
mineralocorticoid
LH
LHRH
FSH
calcitonin

Steroids
prednisone
triamcinolone
hydrocortisone
dexamethasone
betamethosone
prednisolone

Antihypertensives
apresoline
atenolol

Antidiabetic
Diabenese
insulin

Antihistamines
pyribenzamine
chlorpheniramine
diphenhydramine

Antiparasitic
praziquantel
metronidazole
pentamidine

Anticancer
azathioprine
bleomycin
cyclophosphamide
adriamycin
daunorubicin
vincristine
methotrexate
6-TG
6-MP
vinblastine
VP-16
VM-26
cisplatine
FU

Sedatives & Analgesic
morphine
dilaudid
codeine
codeine-like synthetics
demerol
oxymorphone
phenobarbital
barbiturates

Antibiotic
penicillin
tetracycline
erythromycin
cephalothin
imipenem
cefofaxime
carbenicillin

Immunoptherapies
interferon
interleukin-2
monoclonal antibodies
gammaglobulin

Vaccines
influenza
respiratory syncytial virus
Hemophilus influenza vaccine

Antibiotic (continued)
vancomycin
gentamycin
tobramycin
piperacillin
moxalactam
amoxicillin
ampicillin
cefazolin
cefadroxil
cefoxitin
other aminoglycosides

Antifungal
amphotericin B
myconazole
muramyl dipeptide
clotrimazole

Antihypotension
dopamine
dextroamphetamine

Antiviral
acyclovir and derivatives
Winthrop-51711
ribavirin
rimantadine/amantadine
azidothymidine & derivatives
adenine arabinoside
amidine-type protease inhibitors

Proteins and Glycoproteins
lymphokines
  interleukins - 1, 2, 3, 4, 5, and 6
cytokines
  GM-CSF
  M-CSF
  G-CSF
tumor necrosis factor
inhibin
tumor growth factor
Mullerian inhibitors substance
nerve growth factor
fibroblast growth factor
platelet derived growth factor
coagulation factors (e.g. VIII, IX, VII)
insulin
tissue plasminogen activator
histocompatibility antigen
oncogene products
myelin basic protein
collagen
fibronectin
laminin
other proteins made by recombinant DNA technology

Other
cell surface receptor blockers

Nucleic Acids & Analogs
DNA
RNA
methylphosphonates and analogs

A preferred method of making the heterovesicular vesicle or liposome is illustrated in the drawing to which reference is now made. In step 1 (Figure 1) a first aqueous substance of composition 10 to be encapsulated is added to a

first lipid component 12 in the vial 14. The vial 14 is sealed and in step 2 (Figure 2) is mixed and shaken, such as being attached to the head of a vortex mixer to form the first water in oil emulsion 16 containing the first substance of composition 10 to be encapsulated. In step 3 (Figure 3), a second vial 14a, a second aqueous 10a to be encapsulated is added to a second lipid component 12a, and the vial 14a is sealed and in step 4 (Figure 4) is mixed, such as being attached to the head of a vortex mixer to form a second water-in-oil emulsion 16a containing the substance of composition 10a to be encapsulated.

In step 5 (Figure 5) the first 16 and second 16a water in oil emulsions are added together and mixed, such as by hand to make a "chimeric" emulsion.

In step 6 (Figure 6) a portion of the chimeric emulsion from step 5 is individually added to vials containing a third immiscible aqueous component 18a such as by squirting rapidly through a narrow tip pasteur pipette into two one-dram vials, here shown as one.

In step 7 (Figure 7) vials from step 6 are shaken, such as by a vortex mixer, and in step 8 (Figure 8) the chloroform spherule suspension in each vial is transferred from step 7 and the chloroform is evaporated, such as by a stream of nitrogen gas, thereby providing the heterovesicular liposome that contains a first substance in one or more internal aqueous chambers and a second substance in the remaining internal aqueous chambers within a single liposome.

Preferably, each of the substances to be encapsulated are encapsulated in the presence of a hydrochloride, such as hydrochloric acid, which slows their leakage rate from the liposome or vesicle.

As previously mentioned, any biologically active substance, such as illustrated in Table 1, can be encapsulated separately in chambers of the vesicle or liposome.

The following examples set forth presently preferred methods of encapsulating two substances of different compositions in separate chambers of a vesicle or liposome.

Example 1

Preparation of Dideoxycytidine/Glucose Heterovesicular Liposomes

Step 1: A first aqueous substance (one ml of 20 mg/ml dideoxycytidine solution in water with 0.1 N hydrochloric acid) was added into a one-dram vial containing the first lipid component (9.3 umoles of dioleoyl lecithin, 2.1 umoles of dipalmitoyl phosphatidylglycerol, 15 umoles of cholesterol, 1.8 umoles of triolein and one ml of chloroform).

Step 2: The first vial was sealed and attached to the head of a vortex mixer and shaken at maximum speed for 6 minutes to form the first water-in-oil emulsion.

Step 3: In second vial, the second aqueous substance (one ml of 30 mg/ml glucose solution in water with 0.1 N hydrochloric acid) was added into the second lipid component (which is identical to the first lipid component).

Step 4: The second vial was sealed and attached to the head of a vortex mixer and shaken at maximum speed for 6 minutes to form the second water-in-oil emulsion.

Step 5: 0.5 ml of the first emulsion was added to the second vial and mixed by hand to make a "chimeric" emulsion.

Step 6: Half of the "chimeric" emulsion was individually squirted rapidly through a narrow tip Pasteur pipette into one-dram vials, each containing a third immiscible aqueous component (2.5 ml water, 32 mg/ml glucose, 40 mM free-base lysine.

Step 7: The vials from step 6 were shaken on the vortex mixer for 3 seconds at "5" setting to form solvent spherules containing multiple droplets of the first and second aqueous substances within.

Step 8: The chloroform spherule suspensions in each vials were transferred into the bottom of a 2 L beaker containing 4.5 ml of water, 35 mg/ml glucose, and 22 mM free-base lysine. A stream of nitrogen gas at 7 L/min was flushed through the beaker to evaporate chloroform over 5 minutes at 15 deg. C.

The above example describes a method of making heterovesicular liposomes which separately contain glucose in approximately 5/6 of the internal aqueous chambers and separately contain dideoxycytidine in the remaining 1/6 of the internal aqueous chambers within a single liposome. Heterovesicular liposomes containing dideoxycytidine solution as one aqueous substance and glucose as the second aqueous substance were markedly more stable than non-heterovesicular liposomes.

Example 2

This example is for the synthesis of heterovesicular liposomes containing IL-2 (interleukin-2) and lysine hydrochloride: For each batch of liposomes prepared, one ml of water containing 10 mg/ml HSA (Human serum albumin), 1 ug of IL-2, 200 mM lysine HCl pH 7.13 was added into a one-dram vial containing 9.3 umoles of dioleoyl lecithin, 2.1 umoles of dipalmitoyl phosphatidylglycerol, 15 umoles of cholesterol, and 1.8 umoles of triolein and one ml of chloroform (this is the first water-in-oil emulsion). For the second water-in-oil emulsion, 1 ml of lysine HCl (without IL-2) was added into one-dram vial containing 9.3 umoles of dioleoyl lecithin, 2.1 umoles of dipalmitoyl phosphatidylglycerol, 15 umoles of cholesterol, and 1.87 umoles of triolein and one ml of chloroform. Each of the two vials were individually attached to the head of a vortex mixer and shaken sequentially at the maximum speed for 6 minutes.

0.5 ml of the first water-in-oil emulsion was added to the 2 ml of the second emulsion and mixed to make a "chimeric" water-in-oil emulsion. Half of the "chimeric"

emulsion was individually squirted rapidly through a narrow tip Pasteur pipette into one-dram vials, each containing 2.5 ml of 4% glucose in water and 0.1 ml of lysine free base, 200 mM, and shaken at maximum speed for 3 seconds to form chloroform spherules. The chloroform spherule suspensions were transferred into 250 ml Erlenmeyer flask containing 5 ml of 4% glucose in water and 0.2 ml of lysine free base, 200 mM. A stream of nitrogen gas at 7 L/min was flushed through the flask to evaporate chloroform over 5 minutes at 37 degrees C.

Example 3

This example is for the synthesis of heterovesicular liposomes containing ara-C solution as the first aqueous substance and distilled water as the second aqueous substance. For each batch of liposomes prepared, one ml of water containing 100 mg/ml ara-C, pH 1.1 was added into a one-dram vial containing 9.3 umoles of dioleoyl lecithin, 2.1 umoles of dipalmitoyl phosphatidylglycerol, 15 umoles of cholesterol, and 1.8 umoles of triolein and one ml of chloroform, attached to the head of the vortex mixer and shaken at maximum speed for 6 minutes (this is the first water-in-oil emulsion). For the in situ generation of the second water-in-oil emulsion, 1/2 of the content was removed from the first water-in-oil emulsion, and then 1 ml of distilled water was added into the remaining first water-in-oil emulsion and the one-dram vial was shaken for 10 seconds at maximum speed. This resulted in a "chimeric" water-in-oil emulsion. Half of the "chimeric" emulsion was individually squired rapidly through a narrow tip Pasteur pipette into one-dram vials, each containing 2.0 ml of 4% glucose in water and 0.5 ml of lysine free base, 200 mM, and shaken at maximum speed for 3 seconds to form chloroform spherules. The chloroform spherule suspensions were transferred into 250 ml Erlenmeyer flask containing 4 ml of 4% glucose in water and 0.5 ml of lysine free base, 200 mM. A stream of nitrogen gas at 7 L/min was flushed through the

flask to evaporate chloroform over 5 minutes at 37 degrees C.

Example 4

Synthesis of Heterovesicular Liposomes Containing Granulocyte-Macrophase Colony Stimulating Factor (GM-CSF)

Exactly the same procedure was used as in Example 2 except IL-2 was replaced with 1 ug of GM-CSF.

Example 5

Synthesis of Heterovesicular Liposomes of Various Lipid Composition, and Incorporation of Various Materials into Liposomes

In place of using dioleoyl lecithin, dipalmtoyl phosphatidylglyerol, cholesterol, and triolein (TO), and other amphipathic lipids such as phosphatidyl cholines (PC), cardiolipin (CL), dimyristoyl phosphatidylglycerol (DMPG), phosphatidyl ethanolamines (PE), phosphatidyl serines (PS), dimyristoyl phosphatidic acid (DMPA), and other neutral lipids such as tricaprylin (TC) in various combination can be used with similar results. For example, PC/C/CL/TO in 4.5/4.5/1/1 molar ration; DOPC/C/PS/TO in 4.5/4.5/1/1 molar ratio; PC/C/DPPG/TC in 5/4/1/1 molar ratio; PC/C/PG/TC in 5/4/1/1 molar ratio; PE/C/CL/TO in 4.5/4.5/1/1 molar ratio; and PC/C/DMPA/TO in 4.5/4.5/1/1 molar ratio can all be used. To incorporate other water-soluble materials, such as glucose, sucrose, methotrexate, Ponceau S, simply substitute the desired materials for IL-2 in Example 2. Also, other biologically active substances, such as set forth in Table 1, in suitable doses can be similarly substituted for IL-2 as in Example 2.

Example 6

In this example, the triolein in lipid components of above examples are substituted either singly or in combination by other triglycerides, vegetable oils, animal

fats, tocopherols, tocopherol esters, cholesteryl esthers, or hydrocarbons with good results.

Example 7

To make liposomes smaller than that in the foregoing examples, and with reference to Examples 1 or 2, the mechanical strength or duration of shaking or homogenization in Step 4 of Example 1 or 2 was increased. To make liposomes larger, the mechanical strength or duration of shaking or homogenization in Step 4 of Example 1 or 2 was decreased.

The heterovesicular liposomes can be administered to the patients in the normal manner when it is desirable to provide two separate biologically active compounds to the patient for the particular purpose of treatment desired.

The dosage range appropriate for human use includes the range of 1-6000 mg/m to body surface area. The reason that this range is so large is that for some applications, such as subcutaneous administration, the dose required may be quite small, but for other applications, such as intraperitoneal administration, the dose desired to be used may be absolutely enormous. While doses outside the foregoing dose range may be given, this range encompasses the breadth of use for practically all the biologically active substances.

The multivesicular liposomes may be administered by any desired route; for example, intrathecal, intraperitoneal, subcutaneous, intravenous, intralymphatic, oral and submucosal, under many different kinds of epithelia including the bronchialar epithelia, the gastrointestinal epithelia, the urogenital epithelia, and various mucous membranes of the body, and intramuscular.

When encapsulating more than two substances separately in chambers of a liposome, a third (or fourth) aqueous component containing the third or fourth biologically active substance is formed, mixed to form a third or fourth water in oil emulsion, and then combined

with the first and second emulsions and mixed to form a "chimeric" emulsion containing the three or more biologically active substances. The remainder of the process is the same as described when encapsulating two biologically active compounds or substances.

The present invention, therefore, obtains the objects and ends and has the advantages mentioned as well as others inherent therein.

While examples of the invention have been given for the purpose of disclosure, changes can be made therein which are within the spirit of the invention as defined by the appended claims.

What is claimed is:

Claims

1. A heterovesicular lipid vesicle or liposome having at least two different substances, at least one of which is biologically active, encapsulated in separate chambers of the same liposome.

2. A process for producing a heterovesicular lipid vesicle or liposome having at least two different substances, at least one of which is biologically active, separately encapsulated in aqueous chambers thereof comprising the steps of:

(a) providing a first lipid component dissolved in one or more organic solvents and adding into the said lipid component an immiscible first aqueous component containing a first biologically active substance to be encapsulated;

(b) forming a first water-in-oil emulsion from the first two immiscible components;

(c) providing a second lipid component dissolved in one or more organic solvents and adding into the said lipid component an immiscible second aqueous component containing a second substance to be encapsulated;

(d) forming a second water-in-oil emulsion from the second two immiscible components;

(e) forming a chimeric emulsion by combining the first water-in-oil emulsion and the second water-in-oil emulsion;

(f) transferring and immersing the product of step (e) in a third media that is immiscible with said organic solvents;

(g) dispersing the chimeric emulsion to form solvent spherules containing multiple droplets of the first aqueous component containing the first substance and the second aqueous component containing the second substance; and

(h) evaporating the organic solvents from the solvent spherules to form the heterovesicular liposomes.

3. The process according to Claim 2 wherein the first and second lipid components are a phospholipid or an admixture of several phospholipids.

4. The process according to Claim 2 wherein three or more water-in-oil emulsions containing three or more immiscible aqueous components are combined to form the chimeric emulsion.

5. The process according to Claim 2 wherein the first and second lipid components are identical.

6. The process according to Claim 3 wherein the phospholipids are selected from the group consisting of phosphatidylcholine, cardiolipin, phosphatidylethanolamine, sphingomyelin, lysophosphatidylcholine, phosphatidylserine, phosphatidylinositol, phosphatidylglycerol, and phosphatidic acid.

7. The process according to Claim 3 wherein the one or more of the lipid components contain a lipid with a net negative charge or charges.

8. The process according to Claim 3 wherein at least one of the phospholipids is provided in admixture with cholesterol.

9. The process according to Claim 3 wherein at least one of the phospholipids is provided in admixture with stearylamine.

10. The process according to Claim 2 wherein at least one of the first and second substances is a lipophilic biologically active material.

11. The process according to Claim 2 wherein at least one of the first and second lipid components is a neutral lipid either singly or in combination with a substance selected from the group consisting of triglycerides, vegetable oils, animal fats, tocopherols, tocopherol esters, cholesteryl esters, and hydrocarbons.

12. The process according to Claim 2 wherein the organic solvent is selected from the group consisting of ethers, hydrocarbons, halogenated hydrocarbons, halogenated ethers, esters, and combinations thereof.

13. The process according to Claim 2 wherein the hydrochloride is selected from the group consisting of hydrochloric acid, lysine hydrochloride, histidine hydrochloride and combinations thereof.

14. The process according to Claim 2 wherein the biologically active substance is hydrophilic.

15. The process according to Claim 14 wherein the hydrophilic biologically active substance is selected from the group consisting of interleukin-2, cytosine arabinoside, methotrexate, 5-fluorouracil, cisplatin, floxuridine, melphalan, mercaptopurine, thioguanine, thiotepa, vincristine, vinblastine, streptozocin, leuprolide, interferon, calcitonin, doxorubicin, daunorubicin, mitoxanthrone, amacrine, actinomycin, and bleomycin.

16. The process according to Claim 2 wherein the emulsification of the two components is carried out using

methods selected from the group consisting of mechanical agitation, ultrasonic energy, and nozzle atomization.

17. The process according to Claim 2 wherein the third aqueous component contains at least one acid-neutralizing agent.

18. The process according to Claim 17 wherein the acid-neutralizing agent is selected either singly or in combination from the group consisting of free-base lysine and free-base histidine.

19. The process according to Claim 2 wherein the third aqueous component has an ionic strength less than approximately 0.05.

20. The process according to Claim 17 wherein the third aqueous component is an aqueous solution further containing solutes selected from the group consisting of carbohydrates and aminoacids.

21. The process according to Claim 17 wherein the third aqueous component is an aqueous solution containing solutes selected either singly or in combination from the group consisting of glucose, sucrose, lactose, free-base lysine, and free-base histidine.

22. The process according to Claim 2 wherein the dispersion to form solvent spherules is carried out using methods selected from the group consisting of mechanical agitation, ultrasonic energy, and nozzle atomization.

23. The process according to Claim 2 wherein the evaporation of the organic solvent is provided by passing nitrogen gas over the second aqueous component.

24. The process of Claim 2 where, the biologically active substance to be encapsulated is selected from the group consisting of the compositions of Table 1.

25. Heterovesicular liposomes made according to the method of Claim 2.

26. A heterovesicular liposome containing at least two different substances, at least one of which is biologically active, encapsulated in separate chambers of the same liposome, at least one of the substances encapsulated in the presence of a hydrochloride.

27. The process according to Claim 26 wherein the hydrochloride is selected from the group consisting of hydrochloric acid, lysine hydrochloride, histidine hydrochloride and combinations thereof.

28. A heterovesicular liposome containing at least two substances of different compositions, at least one of which is biologically active, encapsulated in separate chambers of the liposome, at least one of the substances encapsulated in the presence of hydrochloric acid or other acid hydrochlorides and a neutralizing agent.

29. The heterovesicular liposome of Claim 26 where, the biologically active composition is selected from the group consisting of the compositions of Table 1.

30. The heterovesicular liposome of Claim 28 where, the biologically active composition is selected from the group consisting of the compositions of Table 1.

31. A method for the treatment of a patient with two different substances, at least one of which is biologically active, comprising administering said

substances to the patient each encapsulated in separate chambers of a heterovesicular liposome.

32. A method for the treatment of a patient with at least two different substances, at least one of which is a biologically active compound, comprising administering said substances to the patient heterovesicular liposomes encapsulating the substances according to Claims 26, 27, 28, 29, 30 or 31.

33. A heterovesicular lipid vesicle or liposome having at least two different substances, at least one of which is biologically active, encapsulated in separate chambers of the same liposome where,

the biologically active substances are selected from the group consisting of antiarrhythmic, antiasthma, antibiotic, anticancer, antidiabetic, antifungal, antihistamines, antihypertensives, antihypotension, antiparasitic, antiviral, cell surface receptor blockers, cardiac glycosides, hormones, immunoptherapies, nucleic acids and analogs, proteins and glycoproteins, sedatives and analgesic, steroids, tranquilizers, and vaccines.

34. The process of claim 2 where,

the biologically active substances are selected from the group consisting of antiarrhythmic, antiasthma, antibiotic, anticancer, antidiabetic, antifungal, antihistamines, antihypertensives, antihypotension, antiparasitic, antiviral, cell surface receptor blockers, cardiac glycosides, hormones, immunoptherapies, nucleic acids and analogs, proteins and glycoproteins, sedatives and analgesic, steroids, tranquilizers, and vaccines.

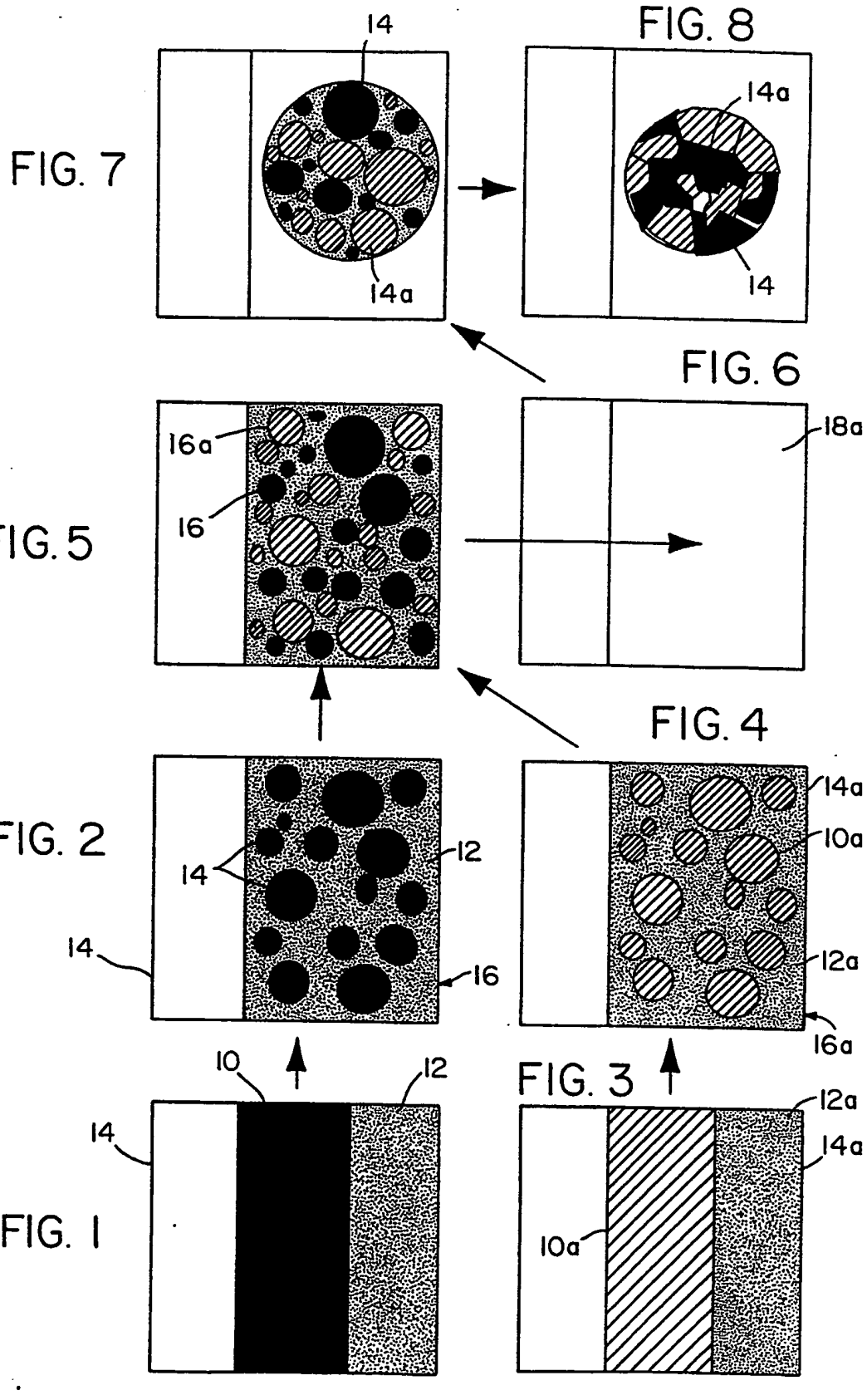
FIG. 7
14
14a
FIG. 8
14a
14
FIG. 6
18a
FIG. 5
16a
16
FIG. 4
14a
10a
12a
16a
FIG. 2
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FIG. 3
12a
14a
10a
FIG. 1
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International Application No PCT/US91/01849

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate [3])

According to International Patent Classification (IPC) or to both National Classification and IPC

IPC(5): A61K 37/22; B01J 13/02
U.S.Cl: 424/450; 264/4.1, 4.3, 4.6

II. FIELDS SEARCHED

Minimum Documentation Searched [4]

Classification System	Classification Symbols
U.S.	424/450; 264/4.1, 4.3, 4.6

Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched [5]

III. DOCUMENTS CONSIDERED TO BE RELEVANT [14]

Category *	Citation of Document, [16] with indication, where appropriate, of the relevant passages [17]	Relevant to Claim No. [18]
Y	Biochimica et Biophysica Acta Vol. 728, 1983 (KIM), Preparation of Multivesicular liposomes, pages 339-348 (See the entire document).	1-34
Y	Chemical Abstracts, Vol. 107, no. 10, issued 1987 (Columbus, Ohio, U.S.A.), Kim, "Multivesicular liposomes containing cytarabine entrapped in the presence of hydrochloric acid for intracavity chemo-therapy" see page 384, col. 1, the abstract No. 83830x,	1-34
Y	WO, A, W085/00515 (The Liposome Company, Inc) 14 February 1985, see abstract and claims	1-34
Y	US, A, 4,235,871 (PAPAHADJOPOULOS) 25 November 1980; See column 4, lines 11-18, column 5, lines 20-22 and examples.	9,11,13,15, 18,21,33 and 34

* Special categories of cited documents: [15]

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search [2]	Date of Mailing of this International Search Report [2]
22 April 1991	24 JUN 1991
International Searching Authority [1]	**Signature of Authorized Officer [20]**
ISA/US	Gollamudi S. Kishore

Form PCT/ISA/210 (second sheet) (May 1986)